

Treatment of mastitis caused by *Staphylococcus aureus* in cows with nanoparticles

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Abstract

One of the most significant zoonotic bacterial diseases is *Staphylococcus aureus* (*S. aureus*), which may infect humans and animals, including dairy cattle worldwide. One of the most issues and financial burdens facing the dairy industry is *S. aureus* caused bovine mastitis, which has a detrimental effect on food safety, production, and animal welfare. Furthermore, it is difficult to cure with conventional medication due to its clever pathophysiology, which includes facultative intracellular parasitism, increasingly severe antibiotic resistance, and biofilm development. Therefore, the creation of nanoparticles is gaining momentum and becoming great instruments for overcoming the treatment challenge associated with *S. aureus* mastitis. In particular, inorganic nanoparticles, polymeric nanoparticles, solid lipid nanoparticles, nanogels, and liposomes seem to be particularly useful. This review focuses on the step-by-step advancement and limitations of nanoparticles in augmenting therapy for *S. aureus* mastitis. The most effective way to prevent new infections is to minimize or eliminate conditions that contribute to the spread of infection and conditions that allow bacteria to contaminate and penetrate the teat canals. First, the challenges of treating *S. aureus* with antimicrobial drugs are examined. Next, *S. aureus* mastitis is controlled. Furthermore, several dietary elements strengthen the animal's defenses against mastitis. Mastitis has decreased when the diet is supplemented with vitamins E and selenium, A and beta carotene, and copper and zinc components that are balanced to fulfill requirements. Second, the benefits of using nanoparticles to treat *S. aureus* mastitis are also outlined. These benefits include enhancing the intracellular penetration and accumulation of the medications that nanoparticles carry, lowering antimicrobial resistance, and avoiding the formation of biofilms. Thirdly, a range of nanoparticle kinds are presented for the purpose of suppressing *S. aureus* mastitis. Lastly, the challenges that still need to be overcome, as well as the potential applications of nanoparticles in the prevention of new infections and the management of *S. aureus* mastitis are covered. The readers will receive sufficient knowledge from this review of the difficulties facing the nanosystem to enable them to create and develop more effective Nano formulations to combat *S. aureus* infections.

Keywords: *Staphylococcus aureus*; bovin mastitis; treatment; nanoparticles

Introduction

One of the most deadly illnesses affecting dairy herds globally is bovine mastitis, which is typically brought on by a variety of bacteria **Ruegg, 2017; Tiwari *et al.*, (2013)**. The most frequent pathogen among these bacteria is *S. aureus*, which is responsible for the most severe cases of mastitis **Monistero *et al.*, (2018)**, and also it cause reproductive issues, a sharp drop in milk revenue, and costs associated with culling infected animals, replacing tainted milk with more expensive veterinary treatment **Botaro *et al.*, (2015)**. *S. aureus* produces toxins in milk that have the potential to result in serious diseases **Johler *et al.*, (2013)**. While **Kaczorektukowska *et al.*, (2022)** showed that Amoxicillin + Clavulonic acid and Gentamycin (100% each) were sensitive to *Staph aureus*, oxytetracycline (98.18%) and Amoxicillin (89.09%) were resistant to *S. aureus*. On antibiotic-resistant *S. aureus* isolated from mastitic milk, Cuo-Nps and Ch-Nps have inhibition zones **Orellano *et al.*, (2021)**. Cuo-Nps and Ch-Nps exhibit antibacterial efficacy against the isolated antibiotic-resistant *S. aureus*, which was isolated from mastitic bovine milk and showed resistance to multiple antibiotics **Hala *et al.*, (2024)**.

Moreover, it has been connected to the subclinical, recurring, and clinical forms of bovine mastitis. Their capacity to remain intracellular result in reinfection **Zhou *et al.*, (2018)**.

The use of antibiotics is one of the main methods for managing mastitis. The treatment benefits are depends on severity of the condition, the medicine of choice, proper drugs use **du Preez, (2000)**. Subsequently, they may revert to a more contagious wild-type phenotype, which increases the likelihood of further infections. Moreover, *S. aureus* develops a greater resistance to antibiotics when they are used often and for extended periods of time **Szweda *et al.*, (2014)**. In the last few years, concerns regarding treatment failure have received a lot of attention. Because of this, scientists have been working nonstop to develop new treatment strategies **Dehkordi *et al.*, (2011)**. New role that nano carriers are playing in the management of cow mastitis and to help researchers understand how to shift their attention to the field of nano carriers in order to discover a

novel approach to combat *S. aureus*-caused mastitis.

S. aureus therapy difficulty

The primary reasons for the treatment's failure are antimicrobial resistance and *S. aureus* ability to remain intracellularly within phagocytes. Numerous variables could be to blame for this, such as a decline in the rate at which routinely used antibiotics are retained in cells, a drop in the pace at which these drugs are absorbed intracellularly, or a decline in the effectiveness of these medications at the lysosomes' acidic pH. Acidic medications may also be ionic at neutral extracellular or cytoplasmic pH, which prevents them from dissolving across the lysosomal membrane. Antibacterial medication's activity is not always present in aqueous solutions due to all of these reasons. Consequently, in order to effectively treat *S. aureus* infections, more specific dose forms are required. If at all possible, these dosage forms should provide the following benefits: (1) broad phagocyte penetration and reserve within cells for an appropriate duration; (2) minimal or nonexistent metabolism within the cells; (3) increased activity against Staphylococci at an acidic pH; (4) distribution through the streak canal. How well antibiotics work to treat mastitis depends on a number of parameters, including their pharmacokinetics—the rate at which they are absorbed and distributed when administered intramarily, their penetration into milk when injected parenterally, and other considerations. **Prescott *et al.* (2000)** state that these properties are related to the kind of vehicle, organic bases given parenterally, even if the former's concentrations ionize in the latterless than those found in blood. Pharmacodynamics is yet another crucial element that must be considered. The antibiotics impede DNA synthesis, stop the formation of bacterial cell walls, and interfere with ribosome activity to stop protein synthesis, among other ways **Normark & Normark, (2002)**. Positive therapy for an infected udder is challenging, if not impossible, because to the following reasons: certain bacteria can generate poisons and enzymes that damage udder tissue and facilitate the entry of germs; survival of

continued undetected by neutrophils and preventing them from being phagocytosed; bacteria's survival and growth within the phagocytes; About half of the *S. aureus* strains that are isolated from sick cattle develop beta-lactamases, microabscesses, and glandular tissue atrophy at the site of infection. All these variables hinder the entry of antibiotics into fibrous membranes.

Therefore, antibiotic resistance in Staphylococci, especially *S. aureus* resistance to penicillin G, is one of the main problems with therapy **Olsen et al., (2006)**. Coagulase-negative staphylococci usually show higher resistance and the capacity to acquire multi-resistance in comparison to *S. aureus* **Pitkälä et al., (2004)**. According to **Haveri et al. (2005)**, some researchers talked about the lack of correlation between the findings of susceptibility testing and the cure rates for mastitis.

Once *S. aureus* attaches itself to host tissues or prosthetic materials, it can spread and persist in a number of ways. Other characteristics of *S. aureus*, such as the production of an anti-phagocytic microcapsule and the creation of an abscess by the zwitterionic capsule, during infection, are essential in assisting the bacteria in avoiding human immunity **O'Riordan & Lee, (2004)**; **Foster, (2005)**. According to **Stephan et al. (2001)**, *Staphylococcus aureus* has the ability to secrete both the extracellular adherent protein and the Staphylococci inhibitory protein, which can hinder neutrophil migration and chemotaxis release to the infection site. Furthermore, *S. aureus* generates leukocidins, which cause leukocytes to be destroyed by opening up cell membrane pores. *S. aureus* produces a variety of enzymes during infection, including lipases, elastases, and proteases, which change the bacteria's ability to assault, damage host tissues, and spread to new locations.

Treatment is made more difficult by *S. aureus* intracellular persistence in macrophages and the mammary epithelium. Furthermore, *S. aureus* can thrive and dwell in unique cell compartments such as the cytosol and endosome, which makes it difficult to remove them from the body and creates a reservoir where infections can recur. In addition, *S.*

aureus small-colony variations (SCVs) are another kind that contributes to recurrent and persistent infection.

SCVs are often resistant to antibiotics and host defense systems after being able to "hide" in host cells in vitro; yet, they are changed to contagious type, which is likely to cause reinfection **Zhou et al., (2018)**.

For instance, it will be challenging to eradicate an infection without removing the prosthetic device if it is infected **Arrecubieta et al., (2006)**. One essential aspect of *S. aureus* pathogenicity is its capacity to create biofilms, and it has been linked to a number of syndromes, including mastitis, because of its capacity to delay phagocytosis, induce persistent antibiotic resistance, and, depending on the disease pattern, either increase or decrease inflammation **Thurlow et al., (2011)**. The dynamic process of biofilm formation makes it possible for planktonic cells to separate and colonize various surfaces after they quickly proliferated. This mechanism has a major impact on encouraging the microbial pathogen to exist in additional infection sites, which leads to the creation of new biofilms and the widespread spread of the infections. Recent research has demonstrated that mice's immune responses to *S. aureus* in biofilm form, which is used to produce acute mastitis in experiments, are more robust than those to **Gogoi-Tiwari et al. (2015)** planktonic cultures **Monistero et al., (2018)**. Recurrence of infection symptoms is caused by altered gene expression of adhesion molecules and toxins, as well as a quick multiplication that came after the separation process.

Thus, the development of biofilms is a significant contributor to the pathophysiology of *S. aureus*, and it is crucial to find alternative therapies that specifically target this component.

Benefits of using nanoparticles to treat *S. aureus* mastitis

It is envisaged that a new dosage form called nanoparticles will be employed intrammary to achieve the desired results.

Drug delivery methods using nanoparticles exhibit a variety of biological and functional characteristics **Garg et al., (2015b; 2015c)**

To address the issues associated with conventional medication, they can be readily modified by adjusting the drug's dosage and

ratio as well as the components that are used in their manufacture, such as polymers, excipients, stabilizers, and others **Garg *et al.*, (2015d)**.

Prevention of the growth of biofilms

Because the nanomaterials prevent the growth of biofilms, they have a substantial impact on the management of *S. aureus* infections. The glycocalyx, which possesses an anionic charge, is the principal constituent of biofilms. This enables it to interact with nanoparticles that are positively charged and able to pierce the dense biofilm **Kulshrestha *et al.*, (2017)**.

The gold nanoparticles dramatically decreased the amount of biofilm that *S. aureus* produced, according to **Sathyanarayanan *et al.* (2013)**. Triclosan, as an antibacterial drug in solution, kills *S. aureus* only outside the biofilm, according to **Liu, (2019)**. However, loading triclosan into a micellar nanocarrier makes it easier for staphylococcal biofilm to penetrate and kill bacteria inside the biofilm.

-Improved parameters for intracellular delivery

Drug's absorption, distribution within cells, and therapeutic effect are all impacted by the different transport pathways. The nanoparticles respond to changes in PH, temperature, redox, and other environmental stimuli by releasing their payload through pores in their membranes. These nanoparticles stay intracellular for extended periods of time. In order to release the payload medicine at the appropriate location, we must synthesize on-demand release nanoparticles during the nanoparticle production process. The size, form, and synthetic chemistry of the nanoparticles, among other physical attributes, influence how well they are transported within cells.

The size of the nanoparticles has a big impact on how they behave in biological fluid and how much of them enter cells. It determines the nanoparticles' stability, toxicity, drug loading and release, and in vivo bio-distribution. The advantages of submicron versus micron size have been extensively studied; in other words, little particles are more effective than large ones at transporting medications to loci that are infected. For example, **Yuan *et al.***

(2017) shown that silver nanoparticles with a size range of 10 nm to 50 nm can be used to effectively cure goat mastitis caused by *S. aureus*. **Chithrani *et al.* (2006)** reported that the maximum cellular absorption of 50 nm-sized silver nanoparticles was observed in adipose-derived stem cells and mammalian cells **Ko *et al.*, (2015)**. Therefore, regulating the size of the nanoparticles is a viable strategy for intracellular drug delivery in the distinct *S. aureus* compartments.

Their form affects the pharmacokinetics of the payload medicine, biological behavior, phagocytosis by macrophages, and cellular uptake of the nanoparticles.

Variations in the charge of the nanoparticles affect the pattern of the endocytosis process. For example, the usage of positive or negative charge particles, such as carboxymethyl chitosan and chitosan hydrochloride, affects the cellular uptake of polymeric nanoparticles. In comparison to negative and neutral charge polymeric nanoparticles, positive charge nanoparticles have a higher rate of phagocytic uptake **He *et al.*, (2010)**. Positive charge nanoparticles are believed to have a nonspecific adherence to normal tissue, making it more difficult for them to reach the diseased area. In contrast, negative charge nanoparticles may be preferred for delivering the treatment deeper into the tissue **Kim *et al.*, (2010)**. **Miao *et al.* (2018)** state that in order to take advantage of the two opposing charges, we can create a type of nanoparticle that carries a positive charge in acidic inflammatory tissue and a negative charge in healthy tissue.

Furthermore, the functionalization of the nanoparticles' surface with "PEG, poloxamer, poloxamine polymers, and other" prevents the particles from being phagocytosed because these polymers make the particles more dispersed and less prone to aggregation, absorption of the protein on their surface, and ionic strength **Moghimi, (1999)**.

Increased resistance to *S. aureus* small-colony variations (SCVs).

Since *S. aureus* infections are hard to treat with antibiotics and to eradicate with the immune system, SCVs have a significant role in the persistence of *S. aureus* infections' resistance

to treatment.

Because of this, a number of studies are attempting to use nanoparticles to increase the effectiveness of antibacterial medications against *S. aureus* SCV phenotypes. Mesoporous silica nanoparticles (MSNP) coated with rifampicin were employed by **Subramaniam et al. (2019)** as a nanocarrier technique to cure intracellular infection brought on by *S. aureus* SCVs. Because MSNP of two sizes, 40 nm and 100 nm, loaded with rifampicin increased the intracellular uptake of rifampicin by RAW 264.7 macrophages infected with *S. aureus* SCVs, more bactericidal action was demonstrated compared with rifampicin alone.

The development of various nanoparticle delivery methods to improve treatment for *S. aureus* infections.

It has been proven by multiple researchers that a variety of nanoparticles, both organic and inorganic, may have been employed in medical studies, particularly for illnesses related to cow mastitis. Nanoparticles can prevent the development of resistance to antibiotics through a variety of mechanisms, including killing bacteria, enhancing the effectiveness of currently available antibiotics by preventing their detection or degradation, and enabling targeted drug delivery to microorganisms so they can use the lowest concentration possible **Wong et al., (2013)**. Moreover, impeding colonization, biofilm formation, and bacterial adherence.

Certain nanocarriers can be conjugated or combined with a variety of antibacterial drugs to enhance their pharmacological activity against susceptible and resistant *S. aureus*. Therefore, medication delivery using nanoparticles is thought to be the best method for eliminating *S. aureus* infections. Additionally, this may lead to effective action against microbes. It is possible to boost both the intracellular concentration of antimicrobial medications and the efficacy of phagocytosis by adding specific macrophage ligands to the nanoparticles **Hua et al., (2014)**.

Liposomes

Nigatu et al. (2018) noted that liposomes may be modified to discharge the drug load at the desired site in reaction to the 39°C temperature of the inflammatory area. Through modifications to the drug's pharmacokinetics and bio-distribution, the liposomes enhance pharmacological efficacy while lowering toxicity. Although the stability of the lipid vesicles is limited due to their decreasing shelf life, they are safe to provide to youngsters as well. Furthermore, as mentioned by **Gabizon et al. (2006)** and **Allen and Martin (2004)**, its manufacturing is intricate and time-consuming.

Polymeric nanoparticles

One of the most effective nanoparticle therapies now being investigated as a potential delivery system for antibiotics is polymeric nanoparticles. Block copolymers, which contain two or more highly hydrophobic polymer chains, are used in the self-assembly process to produce polymeric nanoparticles, which are composed of (biocompatible and biodegradable) polymers.

Moreover, hydrophilic or hydrophobic medicinal compounds as well as macromolecules like peptides, proteins, and nucleic acids have been created to encase them in polymeric nanoparticles **Wang et al., (2012)**. Because of its various benefits, such as its low cost, nontoxicity, biodegradability, and biocompatibility, chitosan is used as a drug delivery vehicle. **Chakraborty et al. (2010)** reported that chitosan nanoparticles loaded with vancomycin and tagged with folic acid improved the vancomycin's transport across bacterial cell membranes, leading to a stronger bactericidal effect against vancomycin-resistant *S. aureus*. Additionally, chitosan nanoparticles loaded with tetracycline stopped *S. aureus* from penetrating cells, while chitosan nanoparticles loaded with *Bacillus natto* stopped biofilm development **Maya et al., (2012)**; **Jiang et al., (2017)**. PLGA (poly-lactic-co-glycolic acid) nanoparticles loaded with gentamycin not only acted as carriers of antibacterial drugs but also showed increased antibacterial efficacy against *S. aureus* by boosting the intracellular accumulation and dispersion of gentamycin **Imbuluzqueta et al.,**

(2010). **Thomas *et al.* (2016)** reported that PLGA nanoparticles loaded with ciprofloxacin and calcium phosphate loaded with levofloxacin and nafcillin sodium inhibited *S. aureus*'s ability to produce biofilms **Bastari *et al.*, (2014)**. Moreover, **Turos *et al.* (2007)** showed that the anti-*S. aureus* infection treatment efficiency of ciprofloxacin was enhanced by glycosylated polyacrylate nanoparticles. Improved drug bioavailability and encapsulation efficiency, controlled payload release, and targeted drug release at the inflammatory and infected areas are further advantages of polymeric nanocarriers **Kumari *et al.*, (2010)**.

On the other hand, reactive groups may influence the stability of the polymer as well as the pace at which the conjugation process progresses **Jijie *et al.*, (2017)**.

SLNs (Solid lipid nanoparticles)

They have drawn interest as a possible therapeutic carrier for intracellular infections since they are biocompatible, biodegradable, and stable **Xie *et al.*, (2011)**. **Xie *et al.*, (2014)**. Another theory is that SLNs can serve as a substitute for liposome and polymeric nanoparticles. For example, studies by **Wang *et al.* (2012)** and **Han *et al.* (2009)** shown how SLNs loaded with tilmicosin could potentially prevent *S. aureus* mastitis. Furthermore, our previous studies **Xie *et al.*, (2017)**; **Li *et al.*, (2019)** demonstrated that docosanoic acid SLNs loaded with enrofloxacin could successfully prolong the period of time that enrofloxacin accumulated and was stored inside the cell. In order to improve enrofloxacin's oral bioavailability, stability, and palatability, we most recently coated the SLNs with an enteric coating. According to studies by **Kalhature *et al.***, vancomycin-loaded SLN had a higher antibacterial activity against *S. aureus* than vancomycin that was free (2014). Furthermore, **Wang *et al.* (2015)** found that SLNs improved the antibacterial efficacy of florfenicol, lauric acid, and retinoic acid against *S. aureus*. **Silva and colleagues, (2015)**. SLNs have several benefits, including as long-term stability, ease of manufacture, reduced toxicity, and the capacity to carry lipophilic and hydrophilic medicines.

Nanogels

Nanogels, which range in size from 20 to 200 nm, are a novel and imaginative type of three-dimensional cross-linked nanocarrier within the field of nanoparticles. These Nano-carriers are used in drug delivery to release pharmaceuticals via several methods, such as photo-isomerization at the target site, thermo-sensitivity, PH responsiveness, and enzyme responsiveness, among others. Compared to other drug carrier systems, nanogels are favored because of their remarkable biocompatibility and essential biodegradability, which make them a very promising option for drug delivery systems **Sultana *et al.*, (2013)**. This prevents the organs from being overly filled with nanogels. Nanogels have no effect on the immune system since they are inert in the bloodstream and internal watery environments (**Rigogliuso *et al.*, (2012)**). Furthermore, nanogels can be administered in a variety of methods, such as "oral, nasal, parenteral, pulmonary, intra-ocular, and topical." As to **Soni and Yadav's (2016)** findings, nanogels are engineered to release the drug at the targeted site in a controlled and uninterrupted fashion with no adverse effects. The activity of biomacromolecules in their natural habitat can be positively enhanced and maintained by the addition of nanogels. One technique to express nanogels is through polymeric micellar nanogels systems. According to **Sultana *et al.* (2013)**, these systems exhibit superior stability across the surfactant micelle, slower patterns of dissociation, and a longer withholding period for loaded medications.

Many different types of nanogels are used to treat mastitis conditions; **Krishna *et al.* (2027)**, for example, confirmed that nano copper gel is effective in treating clinical mastitis. Furthermore, red blood cell (RBC) nanogels have been shown to neutralize

Furthermore, research was done on dextran cross-linked polyacrylamide nanogels loaded with zinc nitrate as an antibacterial agent against MRSA **Malzahn *et al.*, (2014)**. Silver nanoparticle loaded-dextran lysozyme nanogels demonstrated a stronger antibacterial impact against *S. aureus* **Ferrer *et al.*, (2014)**. Additionally, *S. aureus* toxins are decreased by

PLGA nanoparticle-loaded RBC hydrogel **Wang et al., (2015)**, toxins connected to MRSA in the extracellular milieu and promoted macrophage phagocytosis of germs **Zhang et al., (2017)**.

Inorganic metal nanoparticles

Antibacterial and antibiofilm properties can also be attributed to metal nanoparticles. For instance, subclinical mastitis was treated with silver nanoparticles (AgNPs) **Dehkordi et al., (2011)**. They were thought to be an affordable substitute for an extremely expensive antimicrobial solution because of their bactericidal and fungicidal effects via a range of mechanisms, such as cell membrane damage, protein denaturation. The combined effects of antibiotics and AgNPs were also evaluated; for example, erythromycin and AgNPs were used to fight *S. aureus* **Kazemi et al., (2014)**.

Additionally, research has looked at the possible medical applications of selenium, a vital mineral with qualities. To halt the spread of MRSA infections, metal nanoparticles are the subject of extensive research **Hibbitts & O'Leary, (2018)**.

Furthermore, violacein nanoparticles were discovered by **Berni et al. (2013)** to be more potent than common treatments in combating *S. aureus* in the setting of mastitis illness. It was believed at the time that violacein was a common bactericidal agent. **Yang et al.** found that amoxicillin nanoparticles worked just as well against *S. aureus* **(2009)**. Furthermore, according to **Garg and colleagues (2015a)**, lasalocid has demonstrated notable efficacy against mastitis-causing microorganisms other than MRSA and its nano-sized form spreads more swiftly in the udder than its micro-sized form. Because they are easier to create in a range of shapes and forms, have antibacterial qualities, and enhance medication stability, metallic nanoparticles are therefore more beneficial. They do, however, have certain disadvantages, including the tendency to clump together fast, collect in the body after delivery, and release metal ions into the medium and cause cytotoxicity **Jijie et al., (2017)**. Reducing or eliminating the circumstances that cause teat end exposure through infection dissemination is the greatest

strategy to prevent recurrence of *S. aureus* mastitis once it has been treated **Hala et al., (2024)**.

Conclusions

In dairy farming, staphylococcal subclinical mastitis is a complex disease that causes economic losses. The challenges associated with therapy include the quick acquisition of multidrug resistance, the potential for persistent, recurring infections due to biofilm formation, and facultative intracellular parasitism. Because of these, mastitis is a persistent problem and a subject of discussion for numerous research teams.

It is evident that giving too many or inappropriate antibiotics to dairy cows during treatment has a number of negative effects, including raising the risk of antibiotic resistance, lowering antibacterial activity, and lengthening the time needed to check and extend antimicrobial function **Oliver & Murinda, (2012)**. Therefore, the need to overcome the shortcomings of conventional antibiotics is critical. The limitations imposed by antibiotics have been improved recently thanks to developments in nanoparticles with distinct physiochemical characteristics and functionalization **Yah & Simate, (2015)**. In the past few years, a number of distinct nanoparticles have gained popularity for treating Staphylococcal infections. We have provided a quick overview of the most recent research in this field. These nanoparticles exhibit greater intracellular absorption than other conventional drug delivery methods; they also prolong the drug's intracellular accumulation and retention period, enhance its antibacterial activity, lower antimicrobial resistance, and prevent the formation of biofilms. Therefore, broadening our viewpoint to encompass the Nano world can aid in overcoming and addressing the therapy obstacles associated with *S. aureus* mastitis.

Prospects for treating mastitis in the future

We still need to do research and develop new, costly, yet safe Nano formulations that fight *S. aureus* mastitis in order to meet the therapeutic challenges associated with the illness. As was already mentioned, *S. aureus* can infiltrate tissues and reside there in certain

compartments. Therefore, the efficacy of the nanoparticles needs to be improved in order to effectively distribute the medication to the infected region and achieve colocalization between the pharmaceuticals and the intracellular *S. aureus* Xie *et al.*, (2014). To improve tilmicosin's capacity to treat *S. aureus* cow mastitis, our research team combined in-situ hydrogel technology with (SLN) technology to build a self-assembling tilmicosin nanogel Zhou *et al.*, (2019). Additionally, we are developing different nanogels compositions to treat cow mastitis in an effort to advance our research. Further study on stimuli-responsive nanogels is also needed to improve a topical nanogel counter to medical mastitis. Drug administration via nanoparticles has great potential, but there are some obstacles that need to be addressed, like the medication's quick bodily elimination and the loaded drug's premature release before the intended lesion, and the phagocytosis of the immune cells. These problems can be remedied by integrating nanomaterials with natural drug delivery systems, such as bacteria, red blood cells, platelets, and stem cells, which are believed to be smart drug delivery systems and covering nanoparticles with their cell membranes. They allow the medication to build up in the bloodstream and cross cell membranes for a prolonged amount of time in order to stop intracellular infection. Apart from these advantages, stem cells could offer a feasible method for tissue regeneration and increase the effectiveness of treatment for cow mastitis in the future. By neutralizing the bacterial toxin, RBCs can help lessen the harm caused by the bacterial infection. Our groups have recently produced widely applicable protocols for Nano-crystal Nano-suspension and SLNs, which will be beneficial for their use. It is feasible to get nanomaterials from the lab to the clinic, but it requires patience, work, and regulations.

Finally, there has been a lot of interest in creating Nano-robots that might be used for diagnostic and tissue restoration. However, these are cutting-edge research that have never been used before and could be eclipsed by artificial.

References

- Ahangari, Z.; Ghorbanpoor, M. and Shapouri, M. (2017).** Methicillin resistance and selective genetic determinants of *Staphylococcus aureus* isolates with bovine mastitis milk origin. Iran J Microbiol 9:152–9.
- Alhadrami, H. and Al-Hazmi, F. (2017).** Antibacterial activities of titanium oxide nanoparticles. J Bioelectron Nanotechnol 2:5.
- Allen, T.M. and Martin, F.J. (2004).** Advantages of liposomal delivery systems for anthracyclines. Semin Oncol 31:5–15.
- Arrecubieta, C. and Lowy, F.D. (2006).** *Staphylococcus aureus*—eukaryotic cell interactions. In: Fischetti VA, Novick RP, Ferretti JJ, *et al.*, eds. Grampositive pathogens. 2nd ed. Washington (DC): American Society of Microbiology, 517–25.
- Asli, A.; Brouillette, E. and Ster, C. (2017).** Antibiofilm and antibacterial effects of specific chitosan molecules on *Staphylococcus aureus* isolates associated with bovine mastitis. PLoS One 12:e0176988.
- Aswathanarayan, B.J. and Vittal, R.R. (2017).** Antimicrobial, biofilm inhibitory and anti-infective activity of metallic nanoparticles against pathogens MRSA and *Pseudomonas aeruginosa* PA01. J Pharm Nanotechnol 5: 148–53.
- Atulya, M.; Mathew, A.J.; Rao, J.V. and Rao, C.M. (2014).** Influence of milk components in establishing biofilm mediated bacterial mastitis infections in cattle: a fractional factorial approach. J Res Vet Sci 96:25–7.
- Baptista, P.V.; McCusker, M.P. and Carvalho, A. (2018).** Nano-strategies to fight multidrug resistant bacteria—“A Battle of the Titans”. Front Microbiol 9:1441.
- Bastari, K.; Arshath, M. and Ng, Z.H.M. (2014).** A controlled release of antibiotics from calcium phosphate-coated poly (lactico-glycolic acid) particles and their in vitro efficacy against *Staphylococcus aureus* biofilm. J Mater Sci Mater Med 25:747–57.
- Berni, E.; Marcato, P. and Nakazato, G. (2013).** Violacein/poly (ε-caprolactone)/ chitosan nanoparticles against bovine mastitis: antibacterial and ecotoxicity evaluation. J Phys Conf Ser 429:2030.

- Bonventre, P.F. and Gregoriadis, G. (1978).** Killing of intraphagocytic *Staphylococcus aureus* by dihydrostreptomycin entrapped within liposomes. *J Antimicrob Agents Chemother* 13:1049–51.
- Boonkaew, B.; Suwanpreuksa, P. and Cuttle, L. (2014).** Hydrogels containing silver nanoparticles for burn wounds show antimicrobial activity without cytotoxicity. *J Appl Polym Sci* 131:40215.
- Boonyayatra, S.; Rin-ut, S. and Punyapornwithaya, V. (2014).** Association of intramammary infection caused by biofilm-producing pathogens with chronic mastitis in dairy cows. *Int J Dairy Sci* 9:89–95
- Botaro, B.G.; Cortinhas, C.S. and Dibbern, A.G. (2015).** *Staphylococcus aureus* intramammary infection affects milk yield and SCC of dairy cows. *Trop Anim Health Prod* 47:61–6.
- Bozaci, E.; Akar, E. and Ozdogan, E. (2015).** Application of carboxymethylcellulose hydrogel based silver nanocomposites on cotton fabrics for antibacterial property. *J Carbohydr Polym* 134:128–35.
- Breser, M.L.; Felipe, V. and Bohl, L.P. (2018).** Chitosan and cloxacillin combination improve antibiotic efficacy against different lifestyle of coagulase-negative *Staphylococcus* isolates from chronic bovine mastitis. *Sci Rep* 8:5081.
- Brouillette, E. and Malouin, F. (2005).** The pathogenesis and control of *Staphylococcus aureus*-induced mastitis: study models in the mouse. *J Microbes Infect* 7:560–8.
- Brown, A.N.; Smith, K. and Samuels, T.A. (2012).** Nanoparticles functionalized with ampicillin destroy multiple-antibiotic-resistant isolates of *Pseudomonas aeruginosa* and *Enterobacter aerogenes* and methicillin resistant *Staphylococcus aureus*. *Appl Environ Microbiol* 78:2768–74.
- Caiazza, N.C. and O'Toole, G.A. (2003).** Alpha-toxin is required for biofilm formation by *Staphylococcus aureus*. *J. Bacteriol* 185:3214–7.
- Cardozo, V.F.; Lancheros, C.A. and Narciso, A.M. (2014).** Evaluation of antibacterial activity of nitric oxide-releasing polymeric particles against *Staphylococcus aureus* and *Escherichia coli* from bovine mastitis. *Int J Pharm* 473:20–9.
- Chakraborty, S.P.; Sahu, S.K. and Mahapatra, S.K. (2010).** Nanoconjugated vancomycin: new opportunities for the development of anti-VRSA agents. *J Nanotechnol* 21:105103.
- Chithrani, B.D.; Ghazani, A.A. and Chan, W.C. (2006).** Determining the size and shape dependence of gold nanoparticle uptake into mammalian cells. *Nano Lett* 6:662–8.
- Choi, J.B.; Park, J.S. and Khil, M.S. (2013).** Characterization and antimicrobial property of poly (acrylic acid) nanogel containing silver particle prepared by electron beam. *IJMS* 14:11011–23.
- Coxon, A.; Rieu, P. and Barkalow, F. (1996).** A novel role for the beta 2 integrin CD11b/CD18 in neutrophil apoptosis: a homeostatic mechanism in inflammation. *Immunity* 5:653–66.
- Cremieux, A.; Dumitrescu, O. and Lina, G. (2009).** Panton-valentine leukocidins enhances the severity of community-associated methicillin-resistant *Staphylococcus aureus* rabbit osteomyelitis. *PLoS One* 4: e7204.
- Cucarella, C.; Tormo, M.A. and Ubeda, C. (2004).** Role of biofilm-associated protein bap in the pathogenesis of bovine *Staphylococcus aureus*. *Infect Immun* 72:2177–85.
- Cui, F.; Shi, K. and Zhang, L. (2006).** Biodegradable nanoparticles loaded with insulin-phospholipid complex for oral delivery: preparation, in vitro characterization and in vivo evaluation. *J Control Release* 114: 242–50.
- Deb, R.; Kumar, A. and Chakrabort, S. (2013).** Trends in diagnosis and control of bovine mastitis: a review. *Pak J Biol Sci* 16:1653–61.
- Dees, C. and Schultz, R. (1990).** The mechanism of enhanced intraphagocytic killing of bacteria by liposomes containing antibiotics. *J Vet Immunol Immunopathol* 24:135–46.
- Dehkordi, S.H.; Hosseinpour, F. and Kahrizangi, A.E. (2011).** An in vitro evaluation of antibacterial effect of silver nanoparticles on

- Staphylococcus aureus* isolated from bovine subclinical mastitis. *Afr J Biotechnol* 10:10795–7.
- Diep, B.A.; Gill, S.R. and Chang, R.F. (2006).** Complete genome sequence of USA300, an epidemic clone of community-acquired methicillin-resistant *Staphylococcus aureus*. *Lancet* 367:731–9.
- Dinges, M.M.; Orwin, P.M. and Schlievert, P.M. (2000).** Exotoxins of *Staphylococcus aureus*. *Clin Microbiol Rev* 13:16–34.
- Du Preez, J.H. (2000).** Bovine mastitis therapy and why it fails: continuing education. *J S Afr Vet Assoc* 71:201–8.
- Ehrenberg, M.S.; Friedman, A.E. and Finkelstein, J.N. (2009).** The influence of protein adsorption on nanoparticle association with cultured endothelial cells. *J Biomaterials* 30:603–10.
- Esmailou, M.; Zarrini, G.; Ahangarzadeh Rezaee, M. (2017).** Vancomycin capped with silver nanoparticles as an antibacterial agent against multi-drug resistance bacteria. *Adv Pharm Bull* 7:479–83.
- Fernandes, J.B.C.; Zanardo, L.G. and Galvão, N.N. (2011).** *Escherichia coli* from clinical mastitis. *J Vet Diagn Invest* 23:1146–52.
- Ferreira, I.P.; Pilo, E.D.L. and Recio-Despaigne, A.A. (2016).** Bismuth (III) complexes with 2-acetylpyridine- and 2-benzoylpyridine-derived hydrazones: antimicrobial and cytotoxic activities and effects on the clonogenic survival of human solid tumor cells. *Bioorg Med Chem* 24: 2988–98.
- Ferrer, M.C.C.; Dastgheyb, S. and Hickok, N.J. (2014).** Designing nanogel carriers for antibacterial applications. *J Acta Biomater* 10:2105–11.
- Foster, T.J. (2005).** Immune evasion by staphylococci. *Nat Rev Microbiol* 3: 948–58.
- Franci, G.F.A.; Galdiero, S. and Palomba, L. (2015).** Silver nanoparticles as potential antibacterial agents. *Molecules* 20:8856–74.
- Gabizon, A.A.; Shmeeda, H. and Zalipsky, S. (2006).** Pros and cons of the liposome platform in cancer drug targeting. *J Liposome Res* 16:175–83.
- Garg, S.; Wang, W. and Song, Y. (2015a).** Development of intramammary delivery systems containing lasalocid for the treatment of bovine mastitis: impact of solubility improvement on safety, efficacy, and milk distribution in dairy cattle. *Drug Des Devel Ther* 9:631–42.
- Garg, T.; Rath, G. and Goyal, A. (2015b).** Colloidal drug delivery systems: current status and future directions. *Crit Rev Ther Drug Carrier Syst* 32: 89–147.
- Garg, T.; Rath, G. and Goyal, A. (2015c).** Nanotechnological approaches for the effective management of psoriasis. *Artif Cells Nanomed Biotechnol* 10:378–85.
- Garg, T.; Rath, G.; Murthy, R.R. and Gupta, U.D. (2015d).** Current Nano technological approaches for an effective delivery of bioactive drug molecules to overcome drug resistance tuberculosis. *CPD* 21:3076–89.
- Gholipourmalekabadi, M.; Mobaraki, M. and Ghaffari, M. (2017).** Targeted drug delivery based on gold nanoparticle derivatives. *Curr Pharm Des* 23:18–29.
- Gillich, T.; Acikgoz, C. and Isa, L. (2013).** PEG-stabilized core-shell nanoparticles: Impact of linear versus dendritic polymer shell architecture on colloidal properties and the reversibility of temperature-induced aggregation. *J ACS Nano* 7:316–29.
- Gogoi-Tiwari, J. Babra; Waryah, C. and Sunagar, R. (2015).** Typing of *Staphylococcus aureus* isolated from bovine mastitis cases in Australia and India. *Aust Vet J* 93:278–82.
- Gogoi-Tiwari, J.; Williams, V. and Waryah, C.B. (2015).** Comparative studies of the immunogenicity and protective potential of biofilm vs. planktonic *Staphylococcus aureus* vaccine against bovine mastitis using non-invasive mouse mastitis as a model system. *Biofouling* 31:543–54.
- Gomes, F. and Henriques, M. (2016).** Control of bovine mastitis: old and recent therapeutic approaches. *Curr Microbiol* 72:377–82.
- Graf, A.; Rades, T. and Hook, S.M. (2009).** Oral insulin delivery using nanoparticles based on microemulsions with different

- structure-types: optimization and in vivo evaluation. *Eur J Pharm Sci* 37:53–61.
- Guo, G.; Zhou, H. and Wang, Q. (2017).** Nano-layered magnesium fluoride reservoirs on biomaterial surfaces strengthen polymorph nuclear leukocyte resistance to bacterial pathogens. *Nano scale* 9:875–92.
- Gupta, P.V.; Nirwane, A.M.; Belubbi, T. and Nagarsenker, M.S. (2017).** Pulmonary delivery of synergistic combination of fluoroquinolone antibiotic complemented with proteolytic enzyme: a novel antimicrobial and antibiofilm strategy. *J Nanomed Nanotechnol Biol Med* 13:2371–84.
- Hala, A.M. Abdelhady; Nahla, A. Ebied; Eman T. Al-Sokary and Mostafa, S. Abdou (2024).** Evaluation of the effect of nanocopper oxide and nanochitosan on highly antibiotic resistant *Staphylococcus aureus* isolated from normal and mastitic bovine milk. *Egyptian Journal of Animal Health* 4, 2 (2024), 131-147.
- Han, C., Qi, C. and Zhao, B. (2009).** Hydrogenated castor oil nanoparticles as carriers for the subcutaneous administration of tilmicosin: in vitro and in vivo studies. *J Vet Pharm Ther* 32:116–23.
- Haveri, M.; Rosl of, A.; Rantala, L. and Py or al a, S. (2005).** Toxin genes of *Staphylococcus aureus* isolated from bovine intramammary infection of different clinical characteristics and outcome. In Proceedings of 4th IDF International Mastitis Conference. Mastitis in Dairy Production. Current Knowledge and Future Solutions, Maastricht, the Netherlands. Wageningen, the Netherlands: Wageningen Academic Publishers.
- He, C.; Hu, Y. and Yin, L. (2010).** Effects of particle size and surface charge on cellular uptake and biodistribution of polymeric nanoparticles. *J Biomaterials* 31:3657–66.
- Heck, J.G.; Rox, K. and L unsdorf, H. (2018).** Zirconyl clindamycinphosphate antibiotic nanocarriers for targeting intracellular persisting *Staphylococcus aureus*. *ACS Omega* 3:8589–94.
- Hibbitts, A. and O’Leary, C. (2018).** Emerging nanomedicine therapies to counter the rise of methicillin-resistant *Staphylococcus aureus*. *J Mater* 11: 321.
- Hogeveen, H.; Huijps, K. and Lam, T. (2011).** Economic aspects of mastitis: new developments. *N Z Vet J* 59:16–23.
- Hogeveen, H. (2005).** Mastitis is an economic problem. Proceedings of the British Mastitis Conference; 2005; Warwickshire, UK.
- Hsu, C.Y.; Yang, S.C. and Sung, C.T. (2017).** Anti-MRSA malleable liposomes carrying chloramphenicol for ameliorating hair follicle targeting. *IJN* 12:8227–38.
- Hsueh, Y.H.; Tsai, P.H. and Lin, K.S. (2017).** Ph.-dependent antimicrobial properties of copper oxide nanoparticles in *staphylococcus aureus*. *IJMS* 18:793.
- Hua, L.; Hilliard, J. and Shi, Y. (2014).** Assessment of an anti-alpha-toxin monoclonal antibody for prevention and treatment of *Staphylococcus aureus*-induced pneumonia. *J Antimicrob Agents Chemother* 58: 1108–17.
- Huseby, M.J.; Kruse, A.C. and Digre, J. (2010).** Beta toxin catalyzes formation of nucleoprotein matrix in staphylococcal biofilms. *Proc Natl Acad Sci* 107:14407–12.
- Imbuluzqueta, E.; Gamazo, C.; Ariza, J. and Blanco-Prieto, M.J. (2010).** Drug delivery systems for potential treatment of intracellular bacterial infections. *Front Biosci (Landmark Ed)* 15:397–417.
- Jain, V.K.; Singh, M.; Joshi, V.G.; Chhabra, R.; Singh, K. and Rana, Y.S. (2022).** Virulence and antimicrobial resistance gene profiles of *staphylococcus aureus* associated with clinical mastitis in cattle. *Plos One* 17 (5):e0264762.
- Jamaran, S. and Zarif, B.R. (2016).** Synergistic effect of silver nanoparticles with neomycin or gentamicin antibiotics on mastitis-causing *Staphylococcus aureus*. *OJE* 6:452.
- Jiang, X.; Zhou, W. and He, Y. (2017).** Effects of lipopeptide carboxymethyl chitosan nanoparticles on *Staphylococcus aureus* biofilm. *J Biol Reg Homeost Agents* 31:737–43.
- Jijie, R.; Barras, A. and Teodorescu, F. (2017).** Advancements on the molecular design of nanoantibiotics: current level of development and future challenges. *Mol Syst*

- Des Eng 2:349–69.
- Johler, S.; Tichaczek-Dischinger, P.S. and Rau, J. (2013).** Outbreak of Staphylococcal food poisoning due to SEA-producing *Staphylococcus aureus*. Foodborne Pathog Dis 10:777–81.
- Kaczorektukowska, E.; Mataczewska, J.; Sowinska, P. and Szymanska, M. Ewelina (2022).** *Staphylococcus aureus* from subclinical cases of mastitis in dairy cattle in Poland, what are they hiding? Antibiotic resistance and virulence profile. Pathogens 11 (12):1404.
- Kalhpure, R.S.; Mocktar, C. and Sikwal, D.R. (2014).** Ion pairing with linoleic acid simultaneously enhances encapsulation efficiency and antibacterial activity of vancomycin in solid lipid nanoparticles. J Colloids Surf B Biointerfaces 117:303–11.
- Karathanasis, E.; Ayyagari, A.L. and Bhavane, R. (2005).** Preparation of in vivo cleavable agglomerated liposomes suitable for modulated pulmonary drug delivery. J Control Release 103:159–75.
- Kato, N.; Morohoshi, T. and Nozawa, T. (2006).** Control of gram-negative bacterial quorum sensing with cyclodextrin immobilized cellulose ether gel. J Incl Phenom Macrocycl Chem 56:55–9.
- Kaur, T. and Slavcev, R. (2013).** Solid lipid nanoparticles: tuneable anti-cancer gene/drug delivery systems. In: Wei M, Good D, ed. Novel gene therapy approaches. Rijeka (Croatia): Intech Open.
- Kazemi, J.; Ahmadi, M.; Dastmalchi, S.H. and Adibhesami, M. (2014).** Antibacterial effect of silver nanoparticles along with protein synthesis-inhibiting antibiotics on *Staphylococcus aureus* isolated from cattle mastitis. Biol J Micro 2:15–22.
- Kehrli, M.E.; Jr and Harp, J.A. (2001).** Immunity in the mammary gland. J Vet. Clin. North Am Food Anim Pract 17:495–516.
- Kim, B.; Han, G. and Toley, B.J. (2010).** Tuning payload delivery in tumour cyndroids using gold nanoparticles. Nat Nanotech 5:465–72.
- Klement, E.; Chaffer, M. and Leitner, G. (2005).** Assessment of accuracy of disk diffusion tests for the determination of antimicrobial susceptibility of common bovine mastitis pathogens: a novel approach. J Microbial Drug Resist 11:342–50.
- Ko, W.K.; Heo, D.N. and Moon, H.J. (2015).** The effect of gold nanoparticle size on osteogenic differentiation of adipose-derived stem cells. J Colloid Interface Sci 438:68–76.
- Krishna, A.N.; Reddy, M.V.Y.; Reddy, M.C.B. and Padmini, I. (2017).** Formulation, Evaluation of Nano Copper Gel for Treatment of Clinical Mastitis. J Pharm Res 11:554–7.
- Kulshrestha, S.; Qayyum, S. and Khan, A.U. (2017).** Antibiofilm efficacy of green synthesized graphene oxide-silver nano composite using *Lagerstroemia speciosa* floral extract: a comparative study on inhibition of gram positive and gram-negative biofilms. J Microb pathog 103:167–77.
- Kumari, A.; Yadav, S.K. and Yadav, S.C. (2010).** Biodegradable polymeric nanoparticles based drug delivery systems. J Colloids Surf B Biointerfaces 75: 1–18.
- Le Ray, A.M.; Gautier, H. and Laty, M.K. (2005).** In vitro and in vivo bactericidal activities of vancomycin dispersed in porous biodegradable poly (ϵ -caprolactone) microparticles. J Antimicrob Agents Chemother 49: 3025–7.
- Li, C.; Zhou, K. and Chen, D. (2019).** Solid lipid nanoparticles with enteric coating for improving stability, palatability, and oral bioavailability of enrofloxacin. IJN 14:1619–31.
- Li, X.; Robinson, S.M. and Gupta, A. (2014).** Functional gold nanoparticles as potent antimicrobial agents against multi-drug-resistant 1053 bacteria. ACS Nano 8:10682–6.
- Li, Y.; Su, T. and Zhang, Y. (2015).** Liposomal co-delivery of daptomycin and clarithromycin at an optimized ratio for treatment of methicillin-resistant *Staphylococcus aureus* infection. J Drug Deliv 22:627–37.
- Liu, X.; Li, Z. and Wang, X. (2016).** Novel antimicrobial peptide-modified azithromycin-loaded liposomes against methicillin-resistant *Staphylococcus aureus*. IJN 11:6781–94.

- Liu, Y. (2019).** Adaptive antimicrobial nanocarriers for the control of infectious biofilms. [Groningen]: University of Groningen.
- Malzahn, K.; Jamieson, W.D. and Dröge, M. (2014).** Advanced dextran based nanogels for fighting *Staphylococcus aureus* infections by sustained zinc release. *J Mat Chem B* 2:2175–83.
- Matsunaga, T.; Kamata, S.I.; Kakiuchi, N. and Uchida, K. (1993).** Characteristics of *Staphylococcus aureus* isolated from acute, peracute, and chronic bovine mastitis. *J Vet Med Sci* 55:297–300.
- Maya, S.; Indulekha, S. and Sukhithasri, V. (2012).** Efficacy of tetracycline encapsulated O-carboxymethyl chitosan nanoparticles against intracellular infections of *Staphylococcus aureus*. *Int J Biol Macromol* 51: 392–9.
- Mekkawy, A.I.; El-Mokhtar, M.A. and Nafady, N.A. (2017).** In vitro and in vivo evaluation of biologically synthesized silver nanoparticles for topical applications: effect of surface coating and loading into hydrogels. *IJN*. 759: 12-77.
- Menzies, B.E. (2003).** the role of fibronectin binding proteins in the pathogenesis of *Staphylococcus aureus* infections. *Curr Opin Infect Dis* 16: 225–9.
- Miao, Y.; Qiu, Y. and Yang, W. (2018).** Charge reversible and biodegradable Nano carriers showing dual pH-/reduction-sensitive disintegration for rapid site-specific drug delivery. *J Colloids Surf B Biointerfaces* 169:313–20.
- Mocan, L.; Ilie, I. and Matea, C. (2014).** Surface plasmon resonance induced photoactivation of gold nanoparticles as bactericidal agents against methicillin-resistant *Staphylococcus aureus*. *Int J Nanomed* 9: 1453–61.
- Moghimi, S.M. (1999).** Re-establishing the long circulatory behaviour of poloxamine-coated particles after repeated intravenous administration: applications in cancer drug delivery and imaging. *Biochim Biophys Acta* 1472:399–403.
- Mohammed, W.H.; Ali, A.W. and Al-Awady, M.J. (2018).** Evaluation of in vitro drug release kinetics and antibacterial activity of vancomycin HCl loaded nanogel for topical application. *J Pharm Sci Res* 10:2747–56.
- Mohsenabadi, N.; Rajaei, A.; Tabatabaei, M. and Mohsenifar, A. (2018).** Physical and antimicrobial properties of starch-carboxy methyl cellulose film containing rosemary essential oils encapsulated in chitosan nanogel. *Int J Biol Macromol* 112:148–55.
- Monistero, V.; Graber, H. and Pollera, C. (2018).** *Staphylococcus aureus* isolates from bovine mastitis in eight countries: genotypes, detection of genes encoding different toxins and other virulence genes. *Toxins* 10: 247.
- Mukherjee, S.; Ray, S. and Thakur, R. (2009).** Solid lipid nanoparticles: a modern formulation approach in drug delivery system. *Indian J Pharm Sci* 71: 349.
- Nigatu, A.S.; Ashar, H. and Sethuraman, S.N. (2018).** Elastin-like polypeptide incorporated thermally sensitive liposome improve antibiotic therapy against musculoskeletal bacterial pathogens. *Int J Hyperthermia* 34: 201–8.
- Normark, B.H. and Normark, S. (2002).** Evolution and spread of antibiotic resistance. *J Intern Med* 252:91–106.
- O’Riordan, K. and Lee, J.C. (2004).** *Staphylococcus aureus* capsular polysaccharides. *J Clin Microbiol Rev* 17:218–34.
- Oliver, S.P. and Murinda, S.E. (2012).** Antimicrobial resistance of mastitis pathogens. *J Vet Clin Food Anim Pract* 28:165–85.
- Olsen, J.E.; Christensen, H. and Aarestrup, F.M. (2006).** Diversity and evolution of bla_Z from *Staphylococcus aureus* and coagulase-negative staphylococci. *J. Antimicrob Chemother* 57:450–60.
- Orellano, M.S.; Bohl, L.P.; Breser, M.L.; Isaac, P.; Falcone, R.D. and Porporatto, C. (2021).** A comparative study of antimicrobial activity of differently synthesized chitosan nanoparticles against bovine mastitis pathogens. *Soft Matter* 17(3):694-703.
- Ortiz, C.R.; Martinez, T.M. and Lopez, B.J. (2008).** N-acyl-L-homoserine lactones: a class of bacterial quorum-sensing signals alter post-embryonic root development in *Arabidopsis thaliana*. *J Plant Cell Environ* 31:

- 1497–509.
- Otto, M. (2013).** Staphylococcal infections: mechanisms of biofilm maturation and detachment as critical determinants of pathogenicity. *Annu Rev Med* 64:175–88.
- Patidar, A.; Thakur, D.S.; Kumar, P. and Verma, J. (2010).** A review on novel lipid based nanocarriers. *Int J Pharm Pharm Sci* 2:30–5.
- Pattni, B.S.; Chupin, V.V. and Torchilin, V.P. (2015).** New developments in liposomal drug delivery. *Chem Rev* 115:10938–66.
- Petersson-Wolfe, C.M.I. and Jones, G.M. (2010).** Damage caused by *Staphylococcus aureus* mastitis. *VCES* 404:229.
- Pissuwan, D.; Niidome, T. and Cortie, M. (2011).** The forthcoming applications of gold nanoparticles in drug and gene delivery systems. *J Control Rel* 149:65–71.
- Pitkälä, A.; Haveri, M. and Pyörälä, S. (2004).** Bovine mastitis in Finland: 2001 prevalence, distribution of bacteria, and antimicrobial resistance. *J Dairy Sci* 87:2433–41.
- Planchon, S.; Gaillard-Martinie, B. and Dordet-Frisoni, E. (2006).** Formation of biofilm by *Staphylococcus xylosus*. *Int J Food Microbiol* 109:88–96.
- Prescott, J.F.; Baggot, J.D. and Walker, R.D. (2000).** Antimicrobial therapy in veterinary medicine. Editor. Ames, Iowa: Iowa State University Press.
- Prevost, G.; Couppez, P. and Monteil, H. (2003).** *Staphylococcal epidermolysins*. *J Curr Opin Infect Dis* 16:71–6.
- Pumerantz, A.; Muppidi, K. and Agnihotri, S. (2011).** Preparation of liposomal vancomycin and intracellular killing of methicillin-resistant *Staphylococcus aureus* (MRSA). *Int J antimicrob agents* 37:140–4.
- Qasim, M.; Udomluck, N. and Chang, J. (2018).** Antimicrobial activity of silver nanoparticles encapsulated in poly-N-isopropylacrylamide-based polymeric nanoparticles. *IJN* 13:235–49.
- Rainard, P.; Corrales, J. and Barrio, M. (2003).** Leucotoxic activities of *Staphylococcus aureus* strains isolated from cows, ewes, and goats with mastitis: importance of LukM/LukF'-PV leukotoxin. *Clin Diagn Lab Immunol* 10:272–7.
- Reinoso, E.B.; El-Sayed, A. and Lemmler, C. (2008).** Genotyping of *Staphylococcus aureus* isolated from humans, bovine sub-clinical mastitis, and food samples in Argentina. *Microbiol Res* 163:314–22.
- Rigby, K. and DeLeo, F. (2012).** Neutrophils in innate host defense against *Staphylococcus aureus* infections. *Semin Immunopathol* 34:237–59.
- Rigogliuso, S.; Sabatinob, M.A.; Adamoa, G.; Grimaldib, N. and Dispenza, C. (2012).** Nanogels: nanocarriers for drug delivery application. *Chem. Eng Trans* 27:247–52.
- Ruegg, P.L. (2017).** A 100-year review: mastitis detection, management, and prevention. *J Dairy Sci* 100:10381–97.
- Saeb, A.; Alshammari, A.; Al-brahim, H. and Al-rubeaan, K. (2014).** Production of silver nanoparticles with strong and stable antimicrobial activity against highly pathogenic and multidrug resistant bacteria. *Sci World J* 2:704–8.
- Sathyanarayanan, M.B.; Balachandranath, R. and Genji Srinivasulu, Y. (2013).** The effect of gold and iron-oxide nanoparticles on biofilm-forming pathogens. *J ISRN Microbiol* 2013:272086.
- Shi, S.; Jia, J. and Guo, X. (2016).** Reduced *Staphylococcus aureus* biofilm formation in the presence of chitosan-coated iron oxide nanoparticles. *IJN* 11:6499–506.
- Silva, E.L.; Carneiro, G. and de Araujo, L.A. (2015).** Solid lipid nanoparticles loaded with retinoic acid and lauric acid as an alternative for topical treatment of acne vulgaris. *J Nanosci Nanotechnol* 15:792–9.
- Singh, B.N.; Prateeksha, D.K. and Upreti, B.R. (2017).** Bactericidal, quorum quenching and anti-biofilm nanofactories: a new niche for nanotechnologists. *J Crit Rev Biotechnol* 37:525–40.
- Soni, G. and Yadav, K.S. (2016).** Nanogels as potential nanomedicine carrier for treatment of cancer: a mini review of the state of the art. *Saudi Pharm J* 24:133–9.
- Stephan, R.; Annemüller, C.; Hassan, A. and Lemmler, C. (2001).** Characterization of enterotoxigenic *Staphylococcus aureus*

- strains isolated from bovine mastitis in north-east Switzerland. *J Vet Microbiol* 78:373–82.
- Stojkowska, J.; Kosti, C.D. and Jovanovi, C.Z. (2014).** A comprehensive approach to in vitro functional evaluation of Ag/alginate nanocomposite hydrogels. *J Carbohyd Polym* 111:305–14.
- Subramaniam, S.; Thomas, N. and Gustafsson, H. (2019).** Rifampicin-loaded mesoporous silica nanoparticles for the treatment of intracellular infections. *J Antibiotics* 8:39.
- Sultana, F.; Manirujjaman, M.; Imran-Ul-Haque, M.A. and Sharmin, S. (2013).** An overview of nanogel drug delivery system. *J Appl Pharm Sci* 3: 95–105.
- Sun, W.; Wang, Q. and Guo, Y. (2017).** Selenium suppresses inflammation by inducing microRNA-146a in *Staphylococcus aureus*-infected mouse mastitis model. *J Oncotarget* 8:110949.
- Szweda, P.; Schielmann, M. and Frankowska, A. (2014).** Antibiotic resistance in *Staphylococcus aureus* strains isolated from cows with mastitis in the eastern Poland and analysis of susceptibility of resistant strains to alternative non-antibiotic agents: lyso-staphin, nisin and polymyxin B. *J Vet Med Sci* 76:355–62.
- Thomas, N.; Thorn, C. and Richter, K. (2016).** Efficacy of poly-lactic-co-glycolic acid micro- and nanoparticles of ciprofloxacin against bacterial biofilms. *J Pharm Sci* 105:3115–22.
- Thurlow, L.R.; Hanke, M.L. and Fritz, T. (2011).** *Staphylococcus aureus* biofilms prevent macrophage phagocytosis and attenuate inflammation in vivo. *J Immunol* 186:6585–96.
- Tiwari, J.; Babra, C. and Tiwari, H. (2013).** Trends in therapeutic and prevention strategies for management of bovine mastitis: an overview. *J Vaccin Vaccination* 4:1–11.
- Tormo, M.A.; Knecht, E. and G€otz, F. (2005).** Bap-dependent biofilm formation by pathogenic species of *Staphylococcus*: evidence of horizontal gene transfer? *J Microbiol* 151:2465–75.
- Turos, E.; Shim, J.Y. and Wang, Y. (2007).** Antibiotic-conjugated polyacrylate nanoparticles: new opportunities for development of anti-MRSA agents. *J Bioorg Med Chem Lett* 17:53–6.
- Valle, J.; Latasa, C. and Gil, C. (2012).** Bap, a biofilm matrix protein of *Staphylococcus aureus* prevents cellular internalization through binding to GP96 host receptor. *PLoS Pathogens* 8:e1002843.
- Vasudevan, P.; Nair, M.K.M.; Annamalai, T. and Venkitanarayanan, K.S. (2003).** Phenotypic and genotypic characterization of bovine mastitis isolates of *Staphylococcus aureus* for biofilm formation. *J Vet Microbiol* 92: 179–85.
- Vijayakumar, S.; Vinoj, G. and Malaikozhundan, B. (2015).** Plectranthus amboinicus leaf extract mediated synthesis of zinc oxide nanoparticles and its control of methicillin resistant *Staphylococcus aureus* biofilm and blood sucking mosquito larvae. *Spectrochim Acta A Mol Biomol Spectrosc* 137:886–91.
- Wady, A.F.; Machado, A.L. and Foggi, C.C. (2014).** Effect of a silver nanoparticles solution on *Staphylococcus aureus* and *Candida* spp. *J Nanomater* 2014:1–7.
- Wang, A.Z.; Langer, R. and Farokhzad, O.C. (2012).** Nanoparticle delivery of cancer drugs. *Annu Rev Med* 63:185–98.
- Wang, F.; Gao, W. and Thamphiwatana, S. (2015).** Hydrogel retaining toxin-absorbing nanosponges for local treatment of methicillin-Resistant *Staphylococcus aureus* infection. *Adv Mater* 27:3437–43.
- Wang, H.; Bi, C. and Wang, Y. (2018).** J. Selenium ameliorates *Staphylococcus aureus*-induced inflammation in bovine mammary epithelial cells by inhibiting activation of TLR2, NF- κ B and MAPK signaling pathways. *J BMC Vet Res* 14:197.
- Wang, L.; Hu, C. and Shao, L. (2017).** The antimicrobial activity of nanoparticles: present situation and prospects for the future. *IJN* 12:1227–49.
- Wang, T.; Chen, X. and Lu, M. (2015).** Preparation, characterisation and antibacterial activity of a florfenicol-loaded solid lipid nanoparticle suspension. *IET Nanobiotechnol*

- 9:355–61.
- Wang, X.; Zhang, S. and Zhu, L. (2012).** Enhancement of antibacterial activity of tilmicosin against *Staphylococcus aureus* by solid lipid nanoparticles in vitro and in vivo. *Vet J* 191:115–20.
- Wong, I.Y.; Bhatia, S.N. and Toner M. (2013).** Nanotechnology: emerging tools for biology and medicine. *Genes Dev.* 27:2397–408.
- Wu, F.; Meng, G. and He, J. (2014).** Antibiotic-loaded chitosan hydrogel with superior dual functions: antibacterial efficacy and osteoplastic cell responses. *ACS Appl Mater Interfaces* 6:10005–13.
- Xie, S.; Tao, Y. and Pan, Y. (2014).** Biodegradable nanoparticles for intracellular delivery of antimicrobial agents. *J Control Release* 187:101–17.
- Xie, S.; Yang, F. and Tao, Y. (2017).** Enhanced intracellular delivery and antibacterial efficacy of enrofloxacin-loaded docosanoic acid solid lipid nanoparticles against intracellular *Salmonella*. *Sci Rep* 7:41104.
- Xie, S.; Zhu, L. and Dong, Z. (2011).** Preparation, characterization and pharmacokinetics of enrofloxacin-loaded solid lipid nanoparticles: influences of fatty acids. *J Colloids Surf B Biointerfaces* 83:382–7.
- Xiong, M.H.; Li, Y.J. and Bao, Y. (2012).** Bacteria-responsive multifunctional nanogel for targeted antibiotic delivery. *Adv Mater* 24:6175–80.
- Yah, C.S. and Simate, G.S. (2015).** Nanoparticles as potential new generation broad spectrum antimicrobial agents. *Daru* 23:43.
- Yang, X.; Ouyang, W.; Sun, J. and Li, X. (2009).** Post-antibiotic effect of Amoxicillin nanoparticles against main pathogenic bacteria of Bovine mastitis in vitro. *J Northwest A F Univ-Nat Sci Ed* 37:1–6.
- Yuan, Y.G.; Peng, Q.L. and Gurunathan, S. (2017).** Effects of silver nanoparticles on multiple drug-resistant strains of *Staphylococcus aureus* and *Pseudomonas aeruginosa* from mastitis-infected goats: An alternative approach for antimicrobial therapy. *IJMS* 18:569.
- Zecconi, A.; Cesaris, L. and Liandris, E. (2006).** Role of several *Staphylococcus aureus* virulence factors on the inflammatory response in bovine mammary gland. *Microb Pathog* 40:177–83.
- Zhang, Y.; Zhang, J. and Chen, W.P. (2017).** Erythrocyte membrane-coated nanogel for combinatorial antivirulence and responsive antimicrobial delivery against *Staphylococcus aureus* infection. *J Control Release* 263:185–91.
- Zhao, X. and Lacasse, P. (2008).** Mammary tissue damage during bovine mastitis: causes and control. *J Anim Sci* 86:57–65.
- Zhou, K.; Li, C. and Chen, D. (2018).** A review on nanosystem as an effective approach against infections of *Staphylococcus aureus*. *IJN* 13: 7333–47.
- Zhou, K.; Wang, X. and Chen, D. (2019).** Enhanced treatment effects of tilmicosin against *staphylococcus aureus* cow mastitis by self-assembly sodium alginate-chitosan nanogel. *Pharmaceutics* 11: pii: 524.
- Zhou, T.H.; Su, M. and Shang, B.C. (2012).** Nano-hydroxyapatite/b-tricalcium phosphate ceramics scaffolds loaded with cationic liposomal ceftazidime: preparation, release characteristics in vitro and inhibition to *Staphylococcus aureus* biofilms. *J Drug Dev Ind Pharm* 38:1298–304.
- Zollner, T.; Wichelhaus, T. and Hartung, A. (2000).** Colonization with Superantigen-producing *Staphylococcus aureus* is associated with increased severity of atopic dermatitis. *Clin Exp Allergy* 30:994–1000.