Isopod and Vibrio Co-Infections in Fish: Effects and Mitigation Approaches Rehab, Abdel Moneim Qorany; Shimaa, Mohamed Mansour and Walaa Al-Sayed El-Shaer

Department of Fish Diseases, Animal Health Research Institute (AHRI), Agricultural Research Center (ARC)

Review Article

Corresponding author: Rehab Abdel Moneim Qorany E.mail: rehab.qorany@gmail.com

Received in	19/8/2024
Accepted in	24/9/2024

Abstract

Infectious diseases in fish, particularly those involving multiple pathogens, pose a significant threat to wild fish populations and aquaculture industries. Co-infection by Isopod parasites and Vibrio bacteria significantly impacts fish health. This review focuses on these pathogens interactions and combined effects on various fish species. Isopod parasites attach externally to the host, causing physical trauma, stress, and increased vulnerability to secondary infections. In contrast, Vibrio species, known for causing Vibriosis, induce severe Septicemic infections leading to high mortality rates. The co-infection of fish by these pathogens results in compounded negative effects, including elevated morbidity and mortality, posing challenges for disease management and treatment. Our review highlights the complex interplay between isopod parasites and Vibrio bacteria, emphasizing the need for integrated disease control strategies that address the dual threat. Effective monitoring and management practices targeting both parasitic and bacterial infections are essential for improving fish health, aquaculture productivity, and the sustainability of aquatic ecosystems.

Keywords: Isopod, vibrio, co-infections, mitigation, fish

Introduction

In aquatic environments, pathogenic microorganisms significantly threaten economically important fish by causing secondary infections after primary parasitic infestations. Lesions from parasitic infections are often sites for subsequent microbial infections **Ravichandran** *et al.*, (2008). Disease risk in aquaculture is heightened by the presence of pathogens and poor living conditions, with insufficient disease monitoring and climate changes exacerbating the problem Addo *et al.*, (2017). Climate variations can influence parasites directly or indirectly by altering host traits such as distribution, physiology, behavior, and mortality **Lõhmus and Björklund, (2015).**

The order Isopoda, with over 10,300 species found from highland terrestrial environments to deep oceans, belongs to the phylum Arthropoda, subphylum Crustacea Wilson, (2008). Cyclothoid isopods, blood-feeding crustacean parasites, can severely harm fish by infesting their gill chambers or attaching to their surfaces Eissa et al., (2012). Secondary microbial infections typically arise in lesions caused by parasites Ravichandran et al., (2008).Bacterial diseases, such as those caused by Aeromonas, Pseudomonas, and various Vibrio species (Vibrio alginolyticus, Vibrio cholerae, Vibrio vulnificus, and Vibrio anguillarum) Kayansamruaj et al., (2017), pose significant threats to both wild and farmed fish populations Elgendy et al., (2022). Vibriosis is a common disease that affects various species of farmed and wild fish, leading to substantial financial losses **Mohd Nor** *et al.*, (2019).

Isopod Parasites in Fish 1. Overview of Isopod Parasites: Description:

Isopods are small crustaceans, typically ranging from 0.5 to 3.0 cm in length, with a distinctive dorsoventrally flattened body and no carapace **Montelli and Lewis**, (2008). Most parasitic isopods are ectoparasites, but some, like Cryptoniscoidea, are endoparasites of crustacean hosts **Williams and Boyko**, (2012). Manacae, the juvenile form of isopods, have large compound eyes, six pairs of legs (as opposed to seven in juveniles and adults), and strongly setose pleopods that allow them to swim very quickly.

Common Species:

Livoneca redmanii, Nerocila spp., *Cymothoa exigua*, Anilocra spp., and others.

Habitat:

Isopods include both free-living species found in various habitats and parasitic species, mostly affecting fish. Parasitic isopods are generally marine and prefer warmer seas Lester, (2005). In Egypt, they are found in Lake Qarun, Bitter Lakes, Lake Burullus, Lake Manzala, the Red Sea, and the Suez Canal Saied et al., (2024). The main parasitic isopods in marine fish belong to the suborder Cymothoida, specifically the superfamily Cymothooidea. Some families within this group, like Aegidae, Corallanidae, Cymothoidae, and Gnathiidae, are known to parasitize fish at both immature and adult stages, though many are free-living. These isopods attach to the body surface, mouth, gills, or sometimes the nasal cavities of fish. Aegidae differ from Cymothoidae by having less modified percopods, and Corallanidae are typically found in tropical and subtropical regions Wtchariya and Apiruedee, (2020).

Life cycle:

Gravid females use their ventral oostegites to form a brood pouch, or "marsupium," where they lay their eggs. The eggs hatch and undergo two or more moults before reaching the

"manca" or "pullus II" stage. These young are sometimes expelled from the brood pouch simultaneously when the female contracts. The female then molts, feeds, digests food, and prepares to hatch another batch of eggs, often producing multiple batches Williams and Bunkley-Williams, (2019). Some corallanid isopods, like Argathona macronema, parasitize fish, commonly found in the nasal passages of serranids and lutjanids on the Great Barrier Reef Wtchariya and Apiruedee, (2020). Cymothoids damage fish in various ways; their mancae (larval stage) feed aggressively, often killing fry and fingerlings due to the tissue cause Wtchariya damage they and Apiruedee, (2020).

2. Impact on Fish Health:

Cymothoidae isopods can attach to different parts of a fish's body depending on the species, commonly targeting the mouth, gills, fins, and skin Hoffman, (2019). These attachments can cause injuries and deformities, which may impair fish function Ellis et al., (2008) and negatively impact fisheries' welfare and production efficiency Huntingford et al., (2006). Signs of isopod infestation include hypoxia, flaring opercula, and gasping mouths. Permanently attached adult isopods can hinder fish growth and reproduction. Those in the gill chamber can stunt gill development due to attachmentrelated injuries and pressure atrophy, and are often associated with anemia. Isopods in the mouth can alter oral structures, even completely replacing the tongue, as they feed on the host's blood, weakening the fish and making it more susceptible to fatal diseases and secondary bacterial infections (Ghani, 2003).

Isopods exert pressure on the gill lamellae, causing them to shorten, rupture the epithelial layer, release blood, increase mucus production, and impair gas exchange **Helal and Yousef, (2018).** In some instances, isopods attach to the tongue in the buccal cavity, as observed in Carangoides malabaricus, where they firmly anchor to the tongue with their heads facing inward. These isopods occupy the floor of the lower jaw, causing large lesions and tiny pinholes in the tongue where their pereopods piercing claws penetrate host tissues **Rameshkumar and Ravichandran, (2014).** Infested fish gills exhibit varying degrees of degenerative pathological lesions, including destruction, detachment, hyperplasia, and fusion of primary and secondary gill lamellae. Additionally, hyperplasia of epithelial cells in the gill filament **Nadia and Ibrahim**, (2018), lamellar epithelium, and mucous secreting cells, along with lifting of secondary lamellae and congested blood vessels, have been observed **Helal and Yousef**, (2018).

Vibrio Infections in Fish: 1. Overview of Vibrio Infections

Description: Vibrio species are gramnegative, non-spore-forming, rod-shaped bacteria with a single rigid curve. They are motile and have a single polar flagellum when cultured in broth **Noorlis** *et al.*, (2011).

Pathogenic Species: Common pathogenic species of Vibrio affecting fish include *Vibrio harveyi, V. vulnificus, V. alginolyticus,* and *V. parahaemolyticus,* all of which can lead to significant economic losses **Deng** *et al.*, (2020).

Vibriosis in fish:

Vibriosis is a major bacterial disease affecting fish, shellfish, and crustaceans worldwide Mohamed and Abo-Esa, (2007). Outbreaks occur when fish are exposed to infectious agents under stress from chemical, biological, or physical factors Austin and Austin, (2012). In Egypt, Vibriosis is primarily caused by Vibrio species, with V. alginolyticus, V. vulnificus, and V. parahaemolyticus being the most prevalent Gobarah et al., (2022). Vibrio are gramnegative, curved bacteria that are mostly facultative anaerobes, preferring temperatures above 15°C. They are common in aquatic environments, especially in saltwater and brackish water, though V. cholerae can also be found in freshwater Stephen, (2022). Harmful Vibrio species associated with saltwater include V. anguillarum, V. parahaemolyticus, and V. vulnificus, while in freshwater, V. minicus and V. cholerae are more common Fouz et al., (2002).

Pathogenesis:

The development of Vibriosis depends on three main factors: the host, pathogen, and environment. The interaction between the host

(various fish species) and the pathogen (different Vibrio species) greatly influences the severity of the disease. The infection starts with the bacterium binding to the host tissue, a process aided by various virulence factors. The bacteria then rapidly grow and spread to internal organs via the bloodstream. Although the immune system attempts to block the invasion using physical, cellular, and chemical defenses, environmental stress can tip the balance, allowing the disease to establish Manchanayake et al., (2023). Pathogenic Vibrio species possess virulence factors like membrane and secretory proteins, polysaccharide capsules, outer membrane components, siderophores, and biofilm-forming Ina-Salwany et al., (2019).

2. Impact on Fish Health:

The pathology of Vibriosis varies significantly based on the host species, bacterial strain, infection dose, duration, and environmental conditions. Vibriosis, a septicemic illness, is a leading cause of mortality in marine and brackish water fish throughout the year Roberts, (2012). Common clinical signs of Vibrio infection in fish include lethargy, reduced appetite, skin and fin ulcers, and changes in body coloration Mohamad et al., (2019). Primary lesions typically start as skin erosion and ulceration, eventually extending to deeper muscle layers. These lesions are often accompanied by fin erosion, scale loss, exophthalmia, and a swollen abdomen Xie et al., (2020). Naturally infected fish often show multiple hemorrhagic lesions in internal organs like the liver, kidney, intestines, and viscera, along with an increase in the size of visceral organs and ascitesinduced abdominal swelling Sumithra et al., (2019).

Liver examinations of fish infected with Vibrio species reveal a congested portal vein, bile pigment presence, hepatocyte necrosis, bile duct infiltration, and mononuclear cell infiltration. The gills show necrosis, blood vessel congestion, mononuclear cell infiltration, and thickened secondary lamellae. The spleen often exhibits significant fibrosis, hemosiderin accumulation, and hemorrhage. The kidneys typically show mononuclear cell infiltration

around renal tubules, severe hemorrhage, congested glomerular capillaries, and degeneration and necrosis of renal tubular epithelium El-Sharaby *et al.*, (2018).

Isopod and Vibrio Co-infection:

A mixed infection occurs when multiple pathogens such as bacteria, parasites, viruses, or fungi infect the same fish either simultaneously or consecutively, leading to a secondary concurrent illness. These infections amplify the impact of pathogens, weaken disease resistance, and increase the risk of fish mortality Abdel-Latif et al., (2020). Fish populations are particularly vulnerable to disease or death when predisposing factors, such as the presence of ectoparasites, are present Moraes and Martins, (2004). Parasitic isopod wounds, for instance, can serve as entry points for microbial infections, allowing opportunistic bacterial and fungal diseases to take hold. The combination of parasite attachment and these microbial diseases severely affects the infected fish's physiology, often leading to death Raja et al., (2014).

Fish with co-infections of bacteria and parasites have higher mortality rates due to the parasites weakening their immunity **Salama and Yousef, (2020).** Ectoparasite infestations can compromise a fish's natural immunity, making it more prone to bacterial and other infections **Kotob** *et al.*, (2017).

Additionally, parasites themselves may act as vectors for other harmful species Horton and Okamura, (2001). Pathogens can penetrate fish epithelial tissues through mechanical injuries caused by parasite invasion and movement Xu et al., (2012). For example, the species Stolephorus commersonii can develop Vibriosis following isopod bites Rajkumar et al., (2007). The association between the isopod N. orbignyi and bacterial infections, such as Vibrio alginolyticus, Aeromonas sobria, and Staphylococcus aureus, has been linked to significant mortality in Tilapia zillii and Solea vulgaris Younes et al., (2016). While little is known about the host-parasite interaction in disease causation, a study by El-kabany et al. (2023) highlighted concurrent isopod infestation and Vibriosis in Tilapia zillii and Solea aegyptiaca and their impact on fish health. High salinity (30–35 ppt), high temperatures, parasite infestations, and mechanical damage are also associated with an increased risk of bacterial infections. In this context, *Vibrio alginolyticus* strains from *Tilapia zillii and Solea aegyptiaca* in Manzala Lake and Qaroun Lake may be transmitted by isopods.

Mitigation and Control Strategies: 1. Environmental Management:

- Quarantine is the first line of defense. Osmotic shock (freshwater or saltwater dips) can sometimes eliminate isopods, and mechanical removal with forceps is occasionally possible **Rameshkumar and Ravichandran, (2014).**

- A gradual decrease in water temperature elicits various responses from free-living isopods. At 26°C, they exhibit increased activity, but as the temperature drops, their activity progressively decreases, reaching complete dormancy at 15°C, where they lie motionless on the tank floor, appearing almost paralyzed **Saied** *et al.*, (2024).

- Maintaining optimal water conditions to reduce stress and pathogen load, along with regular cleaning and disinfection of aquaculture facilities, is essential. Studies suggest that pond design, water exchanges, draining, lime/ dolomite application, and periodic partial harvesting are effective methods for controlling Vibriosis. Variations in salinity and temperature enhance *Vibrio* spp. capabilities **Urquhart** *et al.*, (2016).

- The direct application of natural substances such as herbal extracts, probiotics, prebiotics, immunostimulants, and non-antibiotics in water can manage aquatic animals and improve defense against Vibrio infections without negative side effects. Maintaining high-quality water management through optimal conditions is essential. Filtration techniques using various mechanical and biological processes also provide an effective alternative for controlling aquaculture diseases in natural habitats. Combining previously researched methods, such as water changes, monitoring for fish infections, and using cage systems, is crucial for better outcomes. Faria *et al.*, (2004).

2. Chemical Treatments:

- Antibiotics: Antibiotics are commonly used in aquaculture to treat bacterial infections, with most Vibrio spp. being susceptible. These antibiotics are usually administered in baths or added to feed. Commonly used antibiotics for treating Vibriosis include oxytetracycline, tetracycline, quinolones, nitrofurans, potentiated sulfonamides, trimethoprim, sarafloxacin, flumequine, and oxolinic acid **Bondad-Reantaso** *et al.*, (2023). Antibiotic treatments are generally employed at the onset of Vibriosis for prevention or urgent care Xu *et al.*, (2023).

- Parasiticides: Diflubenzuron is sometimes needed along with organophosphate treatments to control isopod infestations in water. Treating fish without addressing their environment may only provide temporary relief. It's also important to be aware of the risk of secondary infections that can arise from severe isopod infestations **Rameshkumar and Ravichandran**, (2014).

3. Biological Control:

- Cleaner Fish: Introduce species that naturally eliminate isopods.

- Probiotics: Improve fish health and infection resistance. Probiotics are live microbes that benefit the host by changing its microbial balance, enhancing feed value, and boosting immune responses to pathogens **El-Saadony** *et al.*, (2021). Vaccination is a safer and more effective method for preventing and managing Vibriosis in aquaculture compared to antibiotics. It reduces antibiotic use and stimulates a strong immune response **Delphino** *et al.*, (2019).

Conclusion

Isopod parasites and Vibrio bacteria coinfection significantly threatens fish health in aquaculture. Managing this issue effectively involves improving water quality, enforcing biosecurity measures, and using targeted antimicrobial treatments. Regular monitoring and early detection are vital to prevent outbreaks. Ongoing research is needed to develop sustainable strategies to address these co-infections and maintain fish health and productivity.

References

- Abdel-Latif, H.M.R.; Dawood, M.A.O.; Menanteau-Ledouble, S. and El-Matbouli. M. (2020). The nature and consequences of coinfections in tilapia: A review. J. Fish Dis. 43 (6):651-664.
- Addo, S.; Carrias, A.A.; Williams, M.A.; Liles, M.R.; Terhune, J.S. and Davis, D.A. (2017). Effects of *Bacillus subtilis* strains and the prebiotic Previda on growth, immune parameters, and susceptibility to *Aeromonas hydrophila* infection in Nile tilapia, *Oreochromis niloticus*. Aquaculture Research, 48 (9): 4798-4810.
- Austin, B. and Austin, D.A. (2012). Bacterial fish pathogens: diseases of farmed and wild fish. 5th ed. Chichester, UK: Springer/Prazis.
- Bondad-Reantaso, M.G.; MacKinnon, B.; Karunasagar, I.; Fridman, S.; Alday-Sanz, V.; Brun, E.; Le Groumellec, M.; Li, A.; Surachetpong, W. and Karunasagar, I. (2023). Review of alternatives to antibiotic use in aquaculture. Rev. Aquac.1:1–31.
- Delphino, M.K.V.C.; Barone, R.S.C.; Leal, C.A.G.; Figueiredo, H.C.P.; Gardner, I.A. and Gonçalves, V.S.P. (2019). Economic appraisal of vaccination against *Streptoccocus agalactiae* in Nile tilapia farms in Brazil. Prev. Vet. Med.,162:131–135.
- Deng, Y.; Xu, L.; Chen, H.; Liu, S.; Guo, Z.; Cheng, C.; Ma, H. and Feng, J. (2020). Prevalence, virulence genes, and antimicrobial resistance of Vibrio species isolated from diseased marine fish in South China. Sci. Rep., 10, 14329.
- Eissa, I.A.M.; El-Lamie, M. and Zakai, M. (2012). Studies on Crustacean Diseases of Seabass, Morone Labrax, in Suez Canal, Ismailia Governorate. Life Science Journal, 9 (3).
- Elgendy, M.Y.; Shaalan, M.; Abdelsalam, M.; Eissa, A.E.; El-Adawy, M.M. and Seida, A.A. (2022). Antibacterial activity of silver nanoparticles against antibiotic resistant

Aeromonas veronii infections in Nile tilapia, Oreochromis niloticus (L.), in vitro and in vivo assay. Aquacult. Res., 53(3):901–920.

- El-kabany, N.M.; Badawy, M.F.; Laban, S.E. and Ismail, T.F. (2023). Natural Parasitic and Bacterial Coinfection in Some Fish Species in Egypt. Egyptian Journal of Aquatic Biology and Fisheries, 27(1): 319-334.
- Ellis, T.; Oidtmann, B.; St-Hilaire, S.; Turnbull, J.; North, B.; MacIntyre, C.; Nikolaidis, J.; Hoyle, I.; Kestin, S. and Knowles, T. (2008). Fin erosion in farmed fish. In: Branson E (ed) Fish welfare. Blackwell, Oxford, pp 121–149.
- El-Saadony, M.T.; Alagawany, M.; Patra, A.K.; Kar, I.; Tiwari, R.; Dawood, M.A.O.; Dhama, K. and Abdel-Latif, H.M.R. (2021). The functionality of probiotics in aquaculture: An overview. Fish Shellfish Immunol.,117:36–52.
- El-Sharaby, S.M.A.; Abd-Elgaber, M.; Tarabees, R.; Khalil, R.H.; Ali, M.N. and El-Ballal, S. (2018). Bacteriological and histopathological studies on Vibrio species isolated from naturally infected freshwater fish in the delta region, Egypt. Adv. Anim. Vet. Sci. 6(1): 17-26.
- Faria, E.C.; Brown, B.J.T. and Snook, R.D. (2004). Water toxicity monitoring using *Vibrio fischeri:* a method free of interferences from colour and turbidity. J Environ Monit. 6:97–102.
- **Fouz, B.; Alcaide, E. and Barrera, R. (2002).** Susceptibility of Nile Tilapia *(Oreochromis niloticus)* to vibriosis due to *Vibrio vulnificus* biotype 2 (serovar E). Aquaculture, 212(1-4): 21-30.
- Ghani, N. (2003). Isopod parasites of marine fishes of Pakistan. Proc. Pak. Congr. Zool. 23, 217–221.
- Gobarah, A.D.; Helmy, S.M.; Mahfouz, N.B.; Fahmy, H.A. and Abou Zeid, M.A.E.H.M. (2022). Virulence genes and antibiotic resistance profile of Vibrio species

isolated from fish in Egypt. Veterinary research forum: an international quarterly journal, 13(3): 315–321.

- Helal, A.M. and Yousef, O.E. (2018). Infestation Study of *Livoneca redmanii* (Isopoda, Cymothoidae) on *Mugil cephalus* in Lake Qarun. Egypt. Acad. J. Biol. Sci. B Zool.,10 (1):1-17.
- Hoffman, G.L. (2019). Parasites of North American freshwater fishes. Ithaca: Cornell University Press.
- Horton, T. and Okamura B. (2001). Cymothoid isopod parasites in aquaculture: a review and case study of a Turkish sea bass (*Dicentrarchus labrax*) and sea bream (*Sparus auratus*) farm. Diseases of Aquatic Orgaisms, 46: 181-188.
- Huntingford, F.A.; Adams, C.; Braithwaite, V.A.; Kadri, S.; Pottinger, T.G.; Sandoe, P. and Turnbull, J.F. (2006). Current issues in fish welfare. J Fish Biol 68:332–372.
- Ina-Salwany, M.Y.N.; Al-saari, A.M.; Mursidi, F.A.; Mohd-Aris, A.; Amal, M.N.A.; Kasai, H.; Mino, S.; Sawabe, T. and Zamri-Saad, M. (2019). Vibriosis in fish: a review on disease development and prevention. J. Aquat. Anim. Health, 31: 3-22.
- Kayansamruaj, P.; Dong, H.T.; Hirono, I.; Kondo, H. and Senapin, S. (2017). Genome characterization of piscine 'Scale drop and muscle necrosis syndrome'- associated strain of *Vibrio harveyi* focusing on bacterial virulence determinants. J. Applied Microbiol., 124: 652-666.
- Kotob, M.H.; Menanteau-Ledouble, S.; Kumar, G.; Abdelzaher, M. and El-Matbouli, M. (2017). The impact of co-infections on fish: A review. Veterinary Research., 47(1): 98.
- Lester, R.J.G. (2005). Crustacean parasites. In Marine Parasitology; Rohde, K., Ed.; CSIRO Publishing: Melbourne, Australia, pp. 138– 144.

- Lõhmus, M. and Björklund, M. (2015). Climate change: What will it do to fish parasite interactions. Biological Journal of the Linnean Society, 116(2): 397–411.
- Manchanayake, T.; Salleh, A.; Amal, M.N.A.; Yasin, I.S.M. and Zamri-Saad, M. (2023). Pathology and pathogenesis of Vibrio infection in fish: A review. Aquaculture Reports, 28,101459.
- Mohamed, N.M. and Abo-Esa, J.F.K. (2007). Study on some causative agents infection in Red Sea Shrimp, *Penaeus semisul-catus* in summer season. Egypt. J. Aquat. Biol. and Fish., 11 (3): 845-857
- Mohamad, N.; Amal, M.N.A.; Ina Salwany, M.Y.; Saad, M.Z.; Nasruddin, N.S.; Alsaari, N.; Mino, S. and Sawabe, T. (2019). Vibriosis in cultured marine fishes: a review. Aquaculture, 512.
- Mohd Nor, N.; Yazid, S.H.M.; Daud, H.M.; Amal, M.N.A. and Mohamad, N. (2019). Costs of management practices of Asian seabass (Lates calcarifer Bloch, 1790) cage culture in Malaysia using stochastic model that includes uncertainty in mortality. Aquaculture, 510: 347-352.
- Montelli, L. and Lewis, J. (2008). Survey of Biofouling on Australian Navy Ships Crustacea; Isopoda and Amphipoda Caprellidea; Maritime Platforms Division Defence Scienceand Technology Organisation: Australia.
- Moraes, F.R. and Martins, M.L. (2004). Favourable conditions and principal teleostean diseases in intensive fish farming. Especial topics in tropical intensive freshwater fish farming. São Paulo: Tec Art, pp. 343-383.
- Nadia, G. Ali and Ibrahim, M. Aboyadak (2018). Histopathological alterations and condition factor deterioration accompanied by isopod infestation in *Tilapia zilli, Mugil capito* and *Solea aegyptiaca* from Lake Qaroun. Egyptian Journal of Aquatic Research. V 44 (1) :57-63.

- Noorlis, A.; Ghazali, F.M.; Cheah, Y.K.; Tuan Zainazor, T.C.; Ponniah, J. and Tunung, R. (2011). Prevalence and quantification of Vibrio species and *Vibrio parahaemolyticus* in freshwater fish at hypermarket level. International Food Research Journal. 2011 May 1;18(2).
- Raja, K.; Vijaya Kumar, V.; Karthinkeyan, V.; Saravanakumar, A.; Sindhuja, K. and Gopalakrishnan, A. (2014). Occurrence of isopode *Nerocila phaiopleura* infestation on white fin wolf herring *(Chirocentrus nudus)* from southeast coast of India. J parasitol Dis., 38 (2): 205-207.
- **Rameshkumar, G. and Ravichandran, S.** (2014). Problems caused by isopod parasites in commercial fishes J Parasit Dis. 8(1):138 -141.
- Ravichandran, S.; Rajkumar, M. and Nirmala, S. (2008). Histo- pathology of the infestation of parasitic isopod Jorymatartoor in the host fish Ilishamelastoma. Aqu. Biol. Aqu. 9-12.
- Rajkumar, M.; Thavasi, R.; Perumal, P. and Trilles, J. (2007). Parasite induced vibriosis in *Stolephorus commersonii*. Research Journal of Microbiology, 2(12): 972–977.
- Roberts, R.J. (2012). Fish Pathology. Hoboken, N.J.: Wiley-Blackwell Saied, M.;
 Elsaied, H.; Mabrok, M.; Adel Abdelmageed, Aljilaney S. and Derwa, H. (2024).
 Trials for the Control of Isopods Infesting Lake Qarun, Egypt, with the Impact of Environmental Stresses on their Parasitism. Egyptian Journal of Aquatic Biology & Fisheries Zoology Department, Faculty of Science, Ain Shams University, Cairo, Egypt. ISSN 1110 6131 Vol. 28(1): 1799 1810.
- Saied, M.; Hosam Elsaied; Mahmoud Mabrok; Adel Abdelmageed and Hassan Derwa (2024). Bilocus phylogenetic diversity and biogeographic distribution of isopods infesting Egyptian fishes, The Egyptian Journal of Aquatic Research.12.004.

- Salama, S.S.A. and Yousef, N.S.I. (2020). The impact of co-infection of sea lice and its concurrent some bacterial diseases with field treatment trials in some marine cultured fishes. Egyptian Journal of Aquatic Biology and Fisheries, 24(7-Special issue): 363-381.
- Stephen, G. (2022). An update on Vibriosis, the major bacterial disease shrimp farmers face. Responsible Seafood Advocate,1-8.
- Sumithra, T.G.; Reshma, K.J.; Anusree, V.N.; Sayooj, P.; Sharma, S.R.K.; Suja, G.; Amala, P.V.; Joseph, S. and Sanil, N.K. (2019). Pathological investigations of *Vibrio vulnificus* infection in genetically improved farmed tilapia (*Oreochromis niloticus L.*) cultured at a floating cage farm of India. Aquaculture, 511: 734217.
- Urquhart, E.A.; Jones, S.H. and Yu, J.W. (2016). Environmental conditions associated with elevated *Vibrio parahaemolyticus* concentrations in Great Bay Estuary, New Hampshire. PLoS One. 2016;11:e0155018.
- Williams, E.H.Jr. and Bunkley-Williams, L. (2019). Life Cycle and Life History Strategies of Parasitic Crustacea. Parasitic crustacea (3), 179-266
- Williams, J.D. and Boyko, C.B. (2012). The global diversity of parasitic Isopods associated with crustacean hosts (Isopoda: Bopyroidea and Cryptoniscoidea). PLoS ONE 2012, 7.
- Wilson, G.D.F. (2008). Global diversity of Isopod crustaceans (Crustacea; Isopoda) in freshwater. Hydrobiologia, 595, 231–240.
- Wtchariya, P. and Apiruedee, S. (2020). New Records of Fish Parasitic Isopods (Crustacea: Isopoda) from the Gulf of Thailand. Animals. 10(12), 2298
- Xie, J.; Bu, L.; Jin, S.; Wang, X.; Zhao, Q.; Zhou, S. and Xu, Y. (2020). Outbreak of Vibriosis caused by *Vibrio harveyi* and *Vibrio alginolyticus* in farmed seahorse *Hippocampus kuda* in China. Aquaculture, 523: 735168.

- Xu, D.; Shoemaker, C.A. and Klesius, P.H. (2012). *Ichthyophthirius multifiliis* as a potential vector of *Edwardsiella ictaluri* in channel catfish. FEMS Microbiol Lett., 329 (2): 160-167.
- Xu, K.; Wang, Y.; Yang, W.; Cai, H.; Zhang, Y. and Huang, L. (2023). Strategies for Prevention and Control of Vibriosis in Asian Fish Culture. Vaccines, 11(1): 98.
- Younes, A.M.; Noor Eldin, A.I. and Abd Ellatif, M.A. (2016). A contribution of crustacean isopodoa, bacterial infections and physicochemical parameters in mass mortalities among fishes in Lake Qarun. Research Journal of Pharmaceutical, Biological and Chemical Sciences, 7(2): 1906–1911.